



STUDIES ON BILIRUBIN CLEARANCES OF METHANOLIC LEAF OF VELVET BEANS (*Mucuna pruriens*) ON CCl₄ INDUCED ALBINO RATS



J. B. Minari¹ and M. O. Yakubu^{2,*}

¹Department of Cell Biology & Genetics, University of Lagos, Lagos State, Nigeria

²Department of Biological Sciences, Federal University Wukari, Taraba State, Nigeria

*Corresponding author: marayaemmah@yahoo.com or moyakubu@fuwukari.edu.ng

Received: November 16, 2016

Accepted: March 06, 2017

Abstract: This study investigated the bilirubin clearance and the hepatoprotective properties of the methanolic extract of *Mucuna pruriens* leaves in Carbon Tetrachloride (CCl₄) induced albino rats. CCl₄ was injected into the albino rats to induce jaundice; the rats were given oral dose (20, 40 and 60 mg/kg body weight) of methanolic extract of *M. pruriens* simultaneously. The effect of this treatment on liver enzymes, total bilirubin, total albumin, total protein, urea, creatinine and cholesterol were evaluated. The administration of CCl₄ alone significantly (P<0.05) increased activities of serum, ALT, AST, ALP, GGT, total bilirubin, creatinine, cholesterol. The concentration of albumin and total protein were significantly reduced (P<0.05). Simultaneous treatment of CCl₄ injection and oral administration of 20, 40 and 60 mg/kg body weight of the methanolic extract of *M. pruriens* significantly reversed (P<0.05) these changes in the serum. The above results suggest that the extract may probably possess component that has anti jaundice properties.

Keywords: Bilirubin, carbon tetrachloride, hepatoprotective, Jaundice, *Mucuna pruriens*

Introduction

Jaundice is a clinical symptom presenting as a yellowish pigmentation of the skin, the conjunctiva, and other mucous membranes due to elevated serum bilirubin. Bilirubin is the yellow breakdown product of the non-protein haeme ring of haemoglobin, cytochromes, catalase, peroxidase and tryptophan pyrrolase (Dennerly *et al.*, 2001). It is formed during breakdown of senescent erythrocytes in the spleen, bone marrow, and in hepatic Kupffer cells, and is released into the plasma. Concentration of bilirubin in blood plasma is usually below 1.2 mg/mL.

In the plasma, bilirubin is bound to albumin and transported to the liver for further metabolism (Arias *et al.*, 2011). In the liver, unconjugated bilirubin is metabolized into water-soluble conjugated bilirubin. Unconjugated bilirubin is toxic to many cell types, intracellular organelles and physiological processes. Bilirubin inhibits DNA synthesis and ATPase activity of brain mitochondria, and uncouples oxidative phosphorylation (Hansen, 2000; Wennberg, 2000; Dennerly, 2001). Increased rate of bilirubin production, albumin deficiency, low activity of the bilirubin-conjugating enzyme UDP-glucuronyltransferase, and disorders of the liver can increase the concentration of unconjugated bilirubin (Sherlock and Dooley, 2008). When bilirubin in plasma exceeds 2.5 mg/dL, bilirubin diffuses into body tissues, causing jaundice (VanDeursen *et al.*, 2010; Arias *et al.*, 2011). The conjunctiva of the eye is one of the first tissues to change colour as bilirubin levels rise giving the usually white sclera a yellow appearance. Urine is usually dark in colour (Sherlock and Dooley, 2008).

In general, there are various treatments for jaundice, depending on the underlying cause. For instance, in treating pre-hepatic jaundice, the objective is to prevent the rapid breakdown of red blood cells that cause bilirubin levels to build up in the blood. In cases of infections: such as malaria, use of medication for the treatment of the underlying infection, is usually recommended and, for genetic blood disorders, such as sickle cell anaemia or thalassaemia, blood transfusions may be required to replace the red blood cells. Gilbert's syndrome does not usually require treatment because the jaundice associated with the condition is not particularly serious and does not pose a serious threat to health (Scriven *et al.*, 2000). In recent years, the growing demand for herbal products has been known to be a very good source of alternative treatment traded across countries. The

use of herbs dates back to the early man who used herbs in their raw and cooked forms to keep fit. Since then, herbs such as *Mucuna pruriens* and others are known and generally accepted by many nations as the first art of treatment available to man (Baladrin *et al.*, 1985; Yakubu *et al.*, 2016). *Mucuna pruriens*, a tropical legume, is one of the most popular medicinal plant that is traditionally used in India; and a constituent of more than 200 indigenous drug formulations (Sathiyarayanan and Arulmonzhi, 2007; Kavitha, 2014). *M. pruriens* seed is a natural source of the amino acid L-3,4-dihydroxy phenyl alanine (L-DOPA); the direct precursor to the neuro transmitter dopamine which is used widely in treatment of Parkinson's disease (PD). Serotonin, oxitriptan, nicotine, N,N-DMT, and bufotenine are the other chemicals found in *M. pruriens* in the addition to L-DOPA (Spencer *et al.*, 1995; 1996;). According to Ancient Ayurvedic literature, *Mucuna* is used as a potent aphrodisiac (Amin *et al.*, 1996), geriatric tonic and vermifuge. In addition, *Mucuna* is also grown as food crop, ornamental plant, living mulch and green manure crop.

Traditionally, The Yoruba tribe of Nigeria, have these seed for the treatment of various human and veterinary disease such as fever, constipation, menstruation disorder, oedema, tuberculosis and for the management of sickle cell anaemia (Guerranti *et al.*, 2001; Kavitha and Thangamani, 2014). However, there is still limited scientific information on the treatment of jaundice with *M. pruriens*.

In the absence of reliable hepatoprotective drugs in orthodox medicine, a large number of alternative medicines are recommended for the treatment of liver disorders and are very often claimed to offer significant relief. A number of plants have been shown to possess hepatoprotective properties by improving the antioxidant status (Akhtar *et al.*, 1990; Gupta *et al.*, 1997; Ahmed and Beg, 2001; Ajit *et al.*, 2010). An example of such a plant is *M. pruriens*.

In view of *M. pruriens* being a rich source of natural antioxidants and its numerous uses in alternative medicine, there is a need to conduct more studies on the plant. The primary objective of this investigation was to evaluate the possibility of the extracts of *M. pruriens* in enhancing the bilirubin clearance from the serum when its level is elevated. Also to examine the hepatoprotective effect of the crude methanolic extract of *M. pruriens* using carbon tetrachloride induced rats as experimental models.

Materials and Methods

Plant samples

Mucuna pruriens leaves were obtained from Mushin Market Lagos, identified and authenticated at the herbarium section of the Department of Botany, University of Lagos.

Experimental animals

The twenty five Australian wistar albino rats (*Ratus norvegicus*) of both sexes weighing between 120 – 180 g used for this study were obtained from the College of Medicine Animal Care Unit, University of Lagos, Idi-Araba, Nigeria. The rats were maintained on standard laboratory feed and tap water (*ad libitum*). This procedure was performed according to the rules in Nigeria governing the use of laboratory animals (Carlson *et al.*, 2004) as acceptable internationally.

Chemicals and reagents

Carbon tetrachloride (CCl₄) used is a product of Sigma, United Kingdom. The alkaline phosphatase kit is a product of Teco Diagnostic Limited, Canada, Glutamate Oxaloacetate Transaminase and Glutamate Pyruvate Transaminase kits are products of Randox Laboratories Limited, U.K and the Artesunate tablets used is a product of Mekophar Chemical Pharmaceutical Joint-Stock Company Chi Minh City, Vietnam. All other chemicals used were of analytical grade and obtained from BDH London.

Preparation of plant materials

The *M. pruriens* leaves were washed in tap water and rinsed in sterile distilled H₂O. The leaves were sun-dried for about a week, blended to fine powder with an electric blender, weighed and stored in containers at room temperature until when required for analyses. The *M. pruriens* was extracted with cold methanol and hot water. These were prepared using the method as described by Oyagade *et al.* (1999). The residues obtained were reconstituted in 95% methanol at stock concentration of 0.2 g/ml. The extract was then stored in the refrigerator at 4 ± 2°C until when needed (Omojasola, 2004).

Experimental design

The rats were randomly allotted into five groups of five rats each and categorized as follows:

Group A received 1 ml of distilled water and were tagged the **control group**

Group B received 0.5 mg/kg body weight CCl₄ and were tagged the **untreated group**

Group C received 0.5 mg/kg body weight CCl₄ and 20 mg/kg weight of the *M. pruriens* leaves extract simultaneously

Group D received 0.5 mg/kg body weight CCl₄ and 40 mg/kg body weight of the *M. pruriens* leaves extract simultaneously

Group E received 0.5 mg/kg body weight CCl₄ and 60 mg/kg body weight of the *M. pruriens* leaves extract simultaneously.

Administration of all materials was by oral gavage and treatment lasted for seven days.

Phytochemical screening

Phytochemical analysis of methanolic extracts of *M. pruriens* was carried out as described by Odebiyi and Sofowora (1978).

Biochemical studies

The procedure as described by Reitmann and Frankel (1957) was employed for the assay of aspartate aminotransferase (AST), alanine aminotransferase (ALT) and guanine glutamyltransferase (GGT) activities while that of bilirubin and albumin concentration in the serum was done using the method described by Malloy and Evelyn (1937) and Rodkey (1965), respectively. The total protein concentration was estimated in the serum using the method as described by Plummer (1978). The Jaffe's method using the alkaline picrate method was used for determination of serum creatinine while the Beckman Coulter AU system was used for the determination of the serum urea.

Cholesterol

Total cholesterol was determined using a simple colorimetric assay that measures the amount of cholesterol in serum (Shahjahan *et al.*, 2004).

Creatinine: The Jaffe's method using the alkaline picrate method was used for determination of serum creatinine.

Serum Urea: The Beckman Coulter AU system was used for the determination of the serum urea

Calculation: For serum SI units (mmol/L), multiply the results by 0.357

Statistical analysis

The statistical methods used in analyzing the data include mean, standard deviation and T-test for equality of means. The analysis was performed using Microsoft excel software. Mean differences were considered statistically significantly (P<0.05).

Histopathological analysis

Histopathological analysis was carried out as described by Rolls (2008). The tissues were fixed in 10% formalin for 24 h, cut into thin slices with a scalpel blade and placed in embeddings of labeled cassettes. The tissues were cut into slices of about 0.3 - 0.5 cm thickness in order to ensure proper penetration of processing reagents. This process was followed by dehydration with 70% alcohol (1 h), 90% alcohol (1 h), 90% alcohol (2 h) Absolute alcohol 100% (2 h) and Absolute alcohol 100% (3 h). After dehydration, it is essential to treat tissue with a reagent that mixes with both alcohol and paraffin wax which in turn is removed in the process of wax. This process is called cleaning and the reagent used for this is xylene.

Wax impregnation

This process removes the cleaning agent from the tissue and allows it to be permeated by the molten paraffin wax (impregnation reagent) which was subsequently allowed to harden to produce a block from which sections were cut.

Microtomy

Fine sections of the tissue were obtained using a rotary microtome. The cut sections were then detached from the knife with a Carmel hair brush, placed on a glass slide containing 20% alcohol, and then floated out. Floating out involves gently lowering the strip of sections from the slide on to the surface of water which was 5 – 10°C below the melting point of the wax. The section would float, expand slightly and become flatted. The slide was dipped in the water obliquely and used in picking the section onto it after which it was placed on a hot plate for the surrounding wax to melt off the tissue (Rolls, 2008).

Staining

The slides were stained after 20 min according to the haemotoxylin and Eosin Staining Technique as described by Rolls (2008).

Results and Discussion

Table 1 shows the results obtained from the phytochemical screening of the aqueous and methanolic extracts of *M. pruriens* leaves. It was revealed that tannin, terpenoid, cardiac glycoside, phenol, alkaloid and reducing sugar were detected in the aqueous and methanolic extracts while flavonoid, steroid and phlobatanin were detected in the aqueous extract only. Flavonoid, phenol and alkaloid were more abundant in the aqueous extract than in the methanolic extract, excluding tannin which was more abundant in the methanolic extract than in the aqueous extract in Table 2.

Table 1: Qualitative phytochemical screening of aqueous and methanolic extracts of *M. pruriens* leaves

Phytochemical component	Aqueous	Methanolic
Tannin	+	+
Flavonoid	-	+
Phenol	+	+
Phlobatanin	-	+
Cardiac glycoside	+	+
Alkaloid	+	+
Reducing sugar	+	+
Terpenoid	+	+
Steroid	-	+
Anthraquinone	-	-
Saponin	-	-

Key: (+) = Present, (-) = Absent

Table 2: Quantitative phytochemical screening of aqueous and methanolic extracts of *M. pruriens* leaves

Phytochemicals present	Aqueous	Methanolic
Flavonoid (mg/100g)	31.11 ± 0.23	24.66 ± 0.32
Phenol (mg/100g)	63.65 ± 0.16	15.17 ± 0.11
Alkaloid (mg/100g)	180.35 ± 0.18	127.78 ± 0.48
Tannin (mg/100g)	25.88 ± 0.04	50.36 ± 0.13

Table 3: Effects of methanolic extracts of *M. pruriens* leaves on the body weight of the albino rats after 7 days of administration

	Initial weight (g)	Final weight (g)
Group A	155.60 ± 4.20	155.61 ± 4.21
Group B	152.40 ± 3.21	127.80 ± 3.20
Group C	150.71 ± 3.90	159.10 ± 2.80
Group D	156.80 ± 2.90	166.33 ± 3.10
Group E	163.31 ± 3.20	174.31 ± 2.70

Values are means of four replicates ± S.D; Values with different superscripts across a row are significantly different (P<0.05); Group A received orally, 1 ml of distilled water daily for 7 days; Group B received 0.5 mg/kg body weight CCL₄ for 7 days; Group C received 0.5 mg/kg body weight CCL₄ and 20 mg/kg body weight of the *M. pruriens* leaves extract (orally) simultaneously for 7 days; Group D received 0.5 mg/kg body weight CCL₄ and 40 mg/kg body weight of the *M. pruriens* leaves extract (orally) simultaneously for 7 days; Group E received 0.5 mg/kg body weight CCL₄ and 60 mg/kg body weight of the *M. pruriens* leaves extract (orally) simultaneously for 7 days

In Table 3, there was a significant (P<0.05) increase in the final body weight of the rats that were treated with different doses of methanolic extracts (Groups C, D, and E) when compared with the untreated jaundice induced rats (Group B) which experienced reduction in weight. The concentration of the urea and creatinine is shown in Table 4. The concentration level of urea and creatinine in the serum of rats treated with 40, 60 mg/kg (Group D and E) was significantly increased (P<0.05) when compared to group A. The activity of total bilirubin and con bilirubin was significantly reduced (P<0.05) after the 7 days administration of the dose 20, 40 and 60 mg/kg when compare to group A

The Lipid profile of cholesterol, triglycerides, high-density lipoprotein and low-density lipoprotein in the serum of the rats is revealed in Table 5 the concentration level in cholesterol and triglycerides in the serum of the rats treated with 20 and 40 mg/kg (group C and D) was significantly increased (P<0.05) when compared to group A. while the concentration level in HDL and LDL was significantly reduced in treated dose 60 mg/kg when compared to group A. The enzyme activities of AST, ALP and ALT in the serum of the rats are revealed in Table 6. The ALP and ALT in the serum of the rats was significantly reduced (P<0.05) in all the treatment groups (Group C, D and E) when compared to group A after the 7 days administration of the doses 20, 40 and 60 mg/kg, respectively. The activity of AST was significantly increased (P<0.05) after the 7 days administration of the dose 60 mg/kg when compared to group A.

Table 7 shows the concentration of total protein and albumin in the serum of the albino rats. The concentration of the total protein level was significantly reduced (P<0.05) in the group treated with 20 and 60 mg/kg dosage when compared to group A. The concentration level of albumin in group D significantly increased (P<0.05) when compared to group A after the 7 days administration of the methanolic extracts of *M. pruriens* leaves. Table 8 shows the concentration of cation. The concentration level in cation in the serum of the rats treated with 40 and 60 mg/kg was significantly increased (P<0.05) when compared to group A.

Table 4: Effects of administration of the methanolic extracts of *M. pruriens* leaves on the concentration of urea, creatinine, total bilirubin and con bilirubin in the serum of Jaundice induced albino rats

	Group A	Group B	Group C	Group D	Group E
UREA	34.00±4.04	33.00±2.30	31.66±2.60	45.00±1.15*#α	38.33±2.02#αβ
CREAT	1.30±0.35	1.23±0.14	1.13±0.14	1.86±0.08#α	1.70±0.20#α
TBIL	0.93±0.03	0.80±0.05*	0.83±0.08	0.86±0.03*	0.80±0.05*
ConBIL	0.56±0.03	0.43±0.03*	0.53±0.08	0.53±0.03#	0.46±0.06*

*=P<0.05 when compared with group A; #=P<0.05 when compared with group B; α=P<0.05 when compared with group C; β=P<0.05 when compared with group D; Values are means of four replicates ± S.D; Values with different superscripts across a row are significantly different (P<0.05); Group A received orally, 1ml of distilled water daily for 7 days; Group B received 0.5 mg/kg body weight CCL₄ for 7 days; Group C received 0.5mg/kg body weight CCL₄ and 20 mg/kg body weight of the *M. pruriens* Leaves extract (orally) simultaneously for 7 days; Group D received 0.5mg/kg body weight CCL₄ and 40mg/kg body weight of the *M. pruriens*, Leaves extract (orally) simultaneously for 7 days; Group E received 0.5mg/kg body weight CCL₄ and 60mg/kg body weight of the *M. pruriens* leaves extract (orally) simultaneously for 7days.

Table 5: Effects of administration of the methanolic extracts of *M. pruriens* leaves on the concentration of cholesterol, triglycerides, HDL and LDL in the serum of Jaundice induced albino rats

	Group A	Group B	Group C	Group D	Group E
CHOL	95.33±2.90	94.00±2.88	109.33±5.20*#	102.33±7.21	88.33±3.48*#αβ
HDL	29.33±1.45	27.33±1.45	32.67±1.20*#	31.00±2.64#	26.00±1.52*αβ
TG	65.00±7.0	69.00±4.58	78.67±2.40*#	65.33±3.75α	57.00±1.52*#αβ
LDL	51.66±6.91	56.57±3.99	63.81±2.44*#	51.24±2.60#α	45.18±0.91*#αβ

*=P<0.05 when compared with group A; #=P<0.05 when compared with group B; α=P<0.05 when compared with group C; β=P<0.05 when compared with group D; Values are means of four replicates ± S.D; Values with different superscript across a row are significantly different (P<0.05)

Table 6: Effects of administration of the methanolic extracts of *M. pruriens* leaves on the concentration of AST, ALP and ALT in the serum of Jaundice induced albino rats

	Group A	Group B	Group C	Group D	Group E
AST	59.33±4.33	57.00±5.00	63.67±2.40*	63.00±2.00*	66.00±3.78#
ALT	33.33±2.33	31.66±3.71	36.67±1.45	34.33±1.45	38.00±2.08*#
ALP	59.33±0.33	59.00±0.57	58.67±0.66	60.33±0.88 α	59.33±0.33

*= $P < 0.05$ when compared with group A; #= $P < 0.05$ when compared with group B; $\alpha = P < 0.05$ when compared with group C; $\beta = P < 0.05$ when compared with group E; Values are means of four replicates \pm S.D; Values with different superscripts across a row are significantly different ($P < 0.05$)

Table 7: Effects of administration of the methanolic extracts of *M. pruriens* leaves on the concentration of total protein and albumin in the serum of Jaundice Induced Albino Rats

	Group A	Group B	Group C	Group D	Group E
TP	6.00±0.11	6.30±0.15*	5.83±0.08*#	5.93±0.14*#	5.90±0.05#
ALB	4.06±0.12	4.16±0.03	3.97±0.14#	4.13±0.08	4.03±0.08

*= $P < 0.05$ when compared with group A; #= $P < 0.05$ when compared with group B; $\alpha = P < 0.05$ when compared with group C; $\beta = P < 0.05$ when compared with group D; values are means of four replicates \pm S.D; Values with different superscripts across a row are significantly different ($P < 0.05$)

Table 8: Effects of administration of the methanolic extracts of *M. pruriens* leaves on the concentration of cation in the serum of Jaundice induced albino rats

	Group A	Group B	Group C	Group D	Group E
Ca	9.90±0.11	10.40±0.11*	9.23±0.52*#	10.36±0.14* α	10.10±0.15 α

*= $P < 0.05$ when compared with group A; #= $P < 0.05$ when compared with group B; $\alpha = P < 0.05$ when compared with group C; $\beta = P < 0.05$ when compared with group D; Values are means of four replicates \pm S.D; Values with different superscripts across a row are significantly different ($P < 0.05$)

Histological Structures of Liver Tissues of Albino Rats

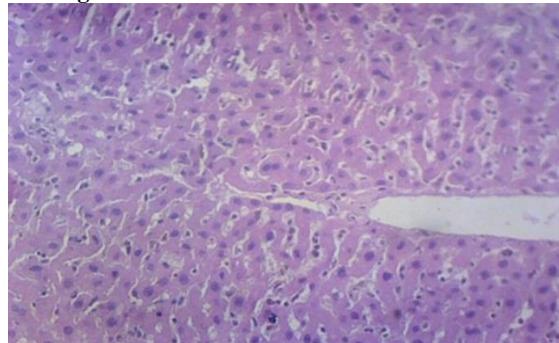


Plate 1: Showing normal hepatocyte (Control group)

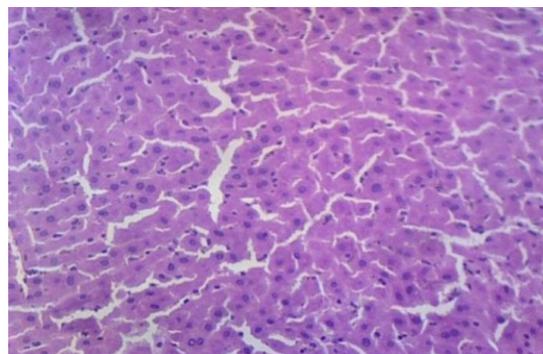


Plate 4: Showing mild deformations in the hepatocytes of the CCL₄ + 20 mg/kg b.wt extract group

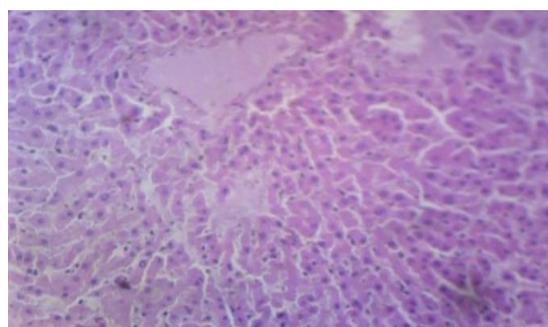


Plate 2: Showing mild deformations in the hepatocytes of the CCL₄ + 60 mg/kg b.wt extract

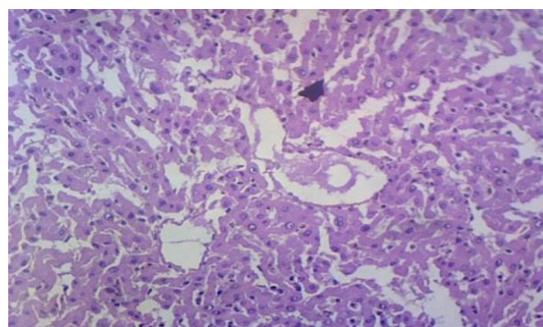


Plate 5: Showing deformations in the hepatocytes of the liver

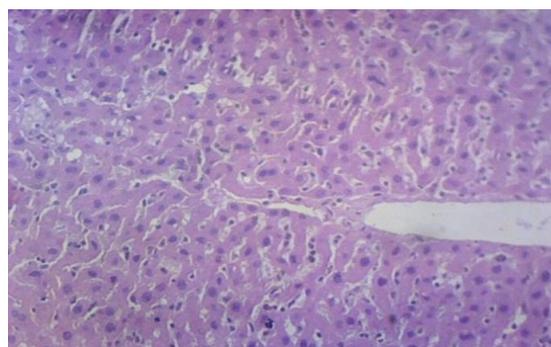


Plate 3: Showing mild deformations in the hepatocytes of the CCL₄ group + 40 mg/kg b.wt extract group

Jaundice is a very important health condition as it is responsible for most liver malfunction, it results from imbalance between bilirubin (a yellowish bile pigment: a reddish yellow bile pigment that is an intermediate product of the breakdown of haemoglobin in the liver) production and excretion. It is the appearance of yellow pigmentation in the skin, sclera and mucous membranes (Dennery *et al.*, 2001; Arias *et al.*, 2011). Under normal circumstances, only a tiny amount of urobilinogen if any is excreted in the urine, but when the liver's function is impaired or when biliary drainage is blocked, some of the conjugated bilirubin leaks out of the hepatocytes, and appears in the urine, turning it dark amber.

The combined increase in the levels of Total cholesterol, Triglyceride and Low density lipoprotein both at low and medium doses (Table 5) is an indication of anti-cardio protective properties of the treatment employed in this study but at the high dose the levels were reduced close to the control levels. These lipids have been shown by various studies to promote/induce the pathogenesis of cardiovascular diseases such as arteriosclerosis, hypertension and heart failure (Vicenova *et al.*, 2009) while there was a corresponding increase in the levels of High density lipoproteins, which is a strong indication of a beneficial consequence (Tackett *et al.*, 2005). This coupled effect points to the fact that the treatment has no negative effect on cardiovascular functions. Increased concentration of low density lipoprotein (LDL) cholesterol or decreased levels of high density lipoprotein (HDL) cholesterol are important risk factors for coronary atherosclerosis. However, an independent association of triglycerides (TG) with atherosclerosis is uncertain (Beatriz *et al.*, 2011). On the other hand, the treatment could have induced systemic hypotension/bradycardia in the animals, which is another index of lowered lipid profile (Seifert *et al.*, 1994; Nermeen *et al.*, 2013). The treatment with high dose resulted in a significant improvement in the lipid profile (Table 5). This finding is similar to previous report by Hamid *et al.* (2011) that the underlying mechanism by which cholesterol is lowered may be due to a decrease in cholesterol absorption from the intestine, by binding with bile acids within the intestine and increasing bile acids excretion (Wu *et al.*, 1999; Binu, 2009) or by decreasing the cholesterol biosynthesis especially by decreasing the 3-hydroxy-3-methyl-glutaryl coenzyme A reductase (HMG CoA reductase) activity, a key enzyme of cholesterol biosynthesis (Sharma *et al.*, 2003) and/or by reducing the NADPH required for fatty acids and cholesterol synthesis (Lorenzetti *et al.*, 1998; Misra and Wagner, 2007).

The significant increase of serum TG level in the low and medium doses is another important finding; recent studies have also shown that triglycerides are independently related to coronary heart disease (Hamid *et al.*, 2011). The observed hyper-triglyceridemic effect may be due to an increase of fatty acids synthesis (Bopanna *et al.*, 1997), enhanced anabolism of LDL, deactivation of LCAT and tissue lipases and/or activation of acetyl-CoA carboxylase (Spina and Cohen, 1988; Lee *et al.*, 2005) and production of triglycerides precursors such as acetyl-CoA and glycerol phosphate. Compared to Silymaril, *Arachis* oil and vitamin E, Methanolic extract of *Argemone Mexican apapavericae* provided a better protective effect as regards the lipid profile.

Also in this study, the levels of alkaline amino transferase, (ALT), AST and ALP of the treated rats which increased marginally when compared to the levels of these enzymes in the control groups indicates that there is a damage in the liver because of increase in the blood. ALT and AST are enzymes involved in amino acid metabolism and used as a marker in liver diseases (Kottai *et al.*, 2010; Ajit and Das, 2010). These enzymes are located in the cell cytoplasm and are emptied into the circulation once the cellular membrane is damaged (Wu *et al.*, 1999). Therefore, the increment of the activities of AST and ALT, in serum may be mainly due to the leakage of these enzymes from the liver cytosol into the blood stream (Navarro *et al.*, 1993), and this is an indication on the hepatotoxic effect of CCl₄.

Aspartate transaminase (AST) is an enzyme that helps metabolize alanine, an amino acid. AST is normally present in blood at low levels. An increase in AST levels may indicate liver damage or disease. The result obtained from (Table 6) indicates that there is an increase in the alanine metabolism in

the liver indicating a hepato-protective effect. Alanine transaminase (ALT) is an enzyme found in the liver that helps the body to metabolize proteins. When the liver is damaged, ALT is released into the bloodstream and levels increase. The graded reduction in the blood levels at the medium and high doses is an indication that the hepatic protein metabolism/synthesis is greatly enhanced by the treatment.

Alkaline phosphatase (ALP) is an enzyme in the liver, bile ducts and bone. Higher than normal levels of ALP may indicate liver damage or disease, such as a blocked bile duct, or certain bone diseases. The dose dependently reduction in the blood levels (Table 6) is an indication that there was no impairment in the biliary system or bone damages which could result in a reduction in red blood cell synthesis. The present study shows exposure to CCl₄ causes an increase in the serum concentrations/values of AST, ALP and ALT. This is similar to a report in another study by Pourgholam *et al.* (2006) they studied the effect of different sub-lethal concentrations of diazinon on grass carp after 45 days and found that levels of ALT, ALP and AST were lower than control.

Similar changes were also observed in *R. frisia* male brood stocks by Luscova *et al.* (2002). They examined the effect of diazinon on carp and showed that Na and K levels were higher and AST, ALP and ALT levels were lower in common carp after being exposed to insecticides. Treatment with medium and high doses resulted in a mild improvement in the liver enzymes status. Goutam *et al.* (2013) has previously shown that there was a significant improvement in the liver enzyme status on the administration following treatment with Methanolic extract of *Argemone Mexican apapavericae*, but this study did not result in a significant improvement. This could be due to the fact that the metabolic function of the liver had been improved following oral treatment with the treatment, the treatment with the ginger oil seems to produce a protective effect on the animal, and this is not only due to the acute nature of the study.

In this study also, plasma samples were analyzed for their bilirubin, urea and creatinine levels. The results obtained in this study also showed that the levels of plasma bilirubin significantly reduced in the treated groups compared to the control (Table 4). The decrease in serum bilirubin (hypobilirubinemia) could have resulted from an increase in liver uptake, conjugation or reduced bilirubin production from haemolysis (Dennery *et al.*, 2001). In addition, the reduction in serum bilirubin indicates liver adaptation as confirmed by the changes in the activities of liver enzymes (AST and ALT). Thus, the reduced level of bilirubin observed in animals in the treated groups when compared with the control group could be attributed to protection over the initial liver damage caused by CCl₄ that the animals are recovering from. The results obtained from this study also indicated that blood urea nitrogen was significantly increased in the treated groups of animals compared to the controls.

From the result obtained, there is a clear indication that the plasma creatinine level was significantly increased at medium and high doses as compared to the control. These findings reveal that there is strong relationship of blood sugar level with creatinine level. The rise in glycemia, involves changes in carbohydrate metabolism and secondarily of lipids and proteins, leading to a loss or degradation of structural proteins due to hepatotoxicity (Wu *et al.*, 1999). Although high levels of creatinine indicate several disturbances in kidney, but also High-serum creatinine level is a marker of muscle wastage. Creatinine (Cr) participates in metabolic reactions within cells and eventually is catabolized in the muscles creating creatinine which is then excreted by the kidney in urine. When Cr is being stored, it is converted to the high energy

form of phosphocreatinine (PCr) which acts as a high-energy reserve in a coupled reaction in which energy derived from donating phosphate is used to regenerate the compound ATP. Since Cr is a critical component of maintaining cellular energy homeostasis, a decrease in creatinine levels, will further contribute to low energy levels in the cells (even in the cells of the brain). The implication of this is that it could serve as a substrate or oxidative stress.

The results from the histological slides helps to buttress the biochemical findings on the hepatotoxicity of CCl₄ and the protective capacity of the graded dose of the extract *M. pruriens* when compared with the standard drug used in this study. This confirmed that the hepatoprotective effect of this extract is only relevant at the high dose of 60 mg/kg body weight.

Conclusion

It can be concluded that *M. pruriens* possesses the phytochemicals that can aid bilirubin clearance from the serum. These phytochemicals also have hepatoprotective properties which enable them to protect the liver against oxidative injuries. It may therefore be recommended as part of the diet for jaundiced individual and also can be used for the management of jaundice in the future.

Acknowledgement

We are grateful to the Department of Cell Biology and Genetics University of Lagos.

Conflict of Interest

The authors declare that there is no conflict of interest.

References

- Ahmad I & Beg AZ 2001. Antimicrobial and Phytochemical studies on 45 Indian medicinal plants against multi-drug resistant human pathogens. *J. Ethnopharm.*, 74: 87-91.
- Ajit, Sarkar & Das AP 2010. Ethnobotanical formulations for the treatment of jaundice by the Mech tribe in Duars of West Bengal. *Indian J. Trade Knowl.*, 9: 134-136.
- Akhtar MS, Qureshi AQ & Iqbal J 1990. Antidiabetic evaluation of *Mucuna pruriens* Linn seeds. *J. Pak. Med. Assoc.*, 40(1): 147-150.
- Amin KMY, Khan, MN, Zillur-Rehman, S & Khan, NA 1996. Sexual function improving effect of *Mucuna pruriens* in sexually normal male rats. *Fitoterapia Milano*, 67(1): 53-56.
- Arias I, Wolkoff A, Boyer J, Shafritz D, Fausto N, Alter H & Cohen D eds. 2011. *The Liver Biology and Pathology*. John Wiley and Sons, New Jersey, p. 1216.
- Baladrin MF, Clocke JA, Wurtele ES & Bolinge WH 1985. *National Plant Chemicals: Source Indu. & Medicinal Materials Sci.*, 228: 1154-1160.
- Beatriz G, Talayero MD, Frank M & Sacks MD 2011. The Role of Triglycerides in Atherosclerosis. *Curr. Cardiol. Rep.*, 13(6): 544-552.
- Bibitha B, Salitha VK, Mohan CV & Valsa AK 2002. Antibacterial activity of different plant extract. Short Communication. *Indian J Microbio.*, 42: 361-363.
- Binu S 2009. Medicinal plants used for treating jaundice (Hepatitis) by the tribals in pathanamthitta district of kerala. *J. Non-timber Forest Products*, 16: 327-330.
- Boer HJ, Kool A, Broberg A, Mziary WR, Hedberg I & Levenfors JJ 2005. *Antifungal & Antibacterial Activity of Some Herbal Remedies from Tanzania*, 96: 461-469.
- Bopanna KN, Kannan J, Sushma G, Balaraman R & Rathod SP 1997. Antidiabetic and antihyperlipaemic effects of neem seed kernel powder on alloxandabetic rabbits. *Indian J. Pharmaco.*, 29(3): 162-167.
- Bravo L, Siddhuraju P & Saura-Calixto F 1998. Effect of various processing methods on the in vitro-starch digestibility and resistant starch content of Indian pulses. *J. Agric. & Food Chem.*, 46(1): 4667-4674.
- Carlson RV, Boyd KM & Webb DJ 2004. The revision of the Declaration of Helsinki: past, present and future. *Br. J. Clin. Pharmacol.*, 57(6): 695-713.
- Dennery PA, Weng YH, Stevenson DK & Yang G. 2001. The biology of bilirubin production. *J. Perinato.*, 21(1): 17-20.
- Gornall AG, Bardawill CJ & David MM 1949. Determination of serum proteins by means of the biuret reaction. *J. Biol. Chem.*, 177: 751-766.
- Goutam B, Dilip G & Rajiv 2013. Argemone Mexicana: Chemical and pharmacological aspects *Rev. Bras. Farmacogn.*, 23: 3
- Guerranti R, Aguiyi JC, Errico E, Pagani R, & Marinello E. 2001. Effects of *Mucuna pruriens* extract on activation of prothrombin by *Echiscarinatus* venom. *J. Ethnopharmac.*, 75(1): 175-180.
- Guerranti R, Ogueli, IG, Bertocci E, Muzzi C, Aguiyi JC, Cianti R, Armini A, Bini L, Leoncini R, Marinello E & Pagani R 2008. Proteomic analysis of the pathophysiological process involved in the anti-snake venom effect of *Mucuna pruriens* extract. *Proteomics*, 8(1): 402-412.
- Gupta M, Mazumder UK, Chakraborti S, Bhattacharya S, Rath N & Bhawal SR 1997. Antiepileptic and anticancer activity of some indigenous plants. *Indian J. Physio. & Allied Sci.*, 51(1): 53-56.
- Hamid O, Schmidt H, Nissan A, Ridolfi L, Aamdal S, Hansson J, Guida M, Hyams DM, Gómez H, Bastholt L, Chasalow SD & Berman D 2011. A prospective phase II trial exploring the association between tumor microenvironment biomarkers and clinical activity of ipilimumab in advanced melanoma. *J. Translational Medicine*, 28(9): 204.
- Hansen TWR 2000. Bilirubin oxidation in brain. *Molecular Genetics & Metabolism*, 71(1): 411-417.
- Kavitha C & Thangamani C 2014. Amazing bean *Mucuna pruriens*: A comprehensive review *J. Medicinal Plants Res.*, 8(2): 138-143.
- Kinard G 2009. Grin taxonomy for plants. *United States Department of Agriculture (USDA) and Agriculture Research Service (ARS)*. Available at: <http://www.ars-grin.gov/cgi-bin/npgs/html/taxgenform.pl>. Accessed 1st May, 2015.
- KottaiMuthu A, Kumar DS, Smith AA & Manavalan R 2010. *In vitro* antioxidant activity of various extracts of whole plant of *Mucuna pruriens* (Linn). *Int. J. PharmTech Res.*, 2(1): 2063-207.
- Lee HY, Bahn SC, Shin JS, Hwang I, Back K, Doelling JH & Ryu SB 2005. Multiple forms of secretory phospholipase A2 in plants. *Progress in Lipid Res.*, 44(1): 52-67.
- Lorenzetti E, MacIsaac S, Arnason JT, Awang DVC & Buckles D 1998. *The phytochemistry, toxicology and food potential of velvet bean (Mucuna aadans spp. Fabaceae)*. In: Buckles, D, Osiname, O, Galiba, M & Galiano, G. Eds. *Cover crops of West Africa: Contributing To Sustainable Agriculture*. IDRC, Ottawa and IITA, Ibadan, p. 318.
- Lusková V, Svoboda M & Kolářová J 2002. The Effect of Diazinon on Blood Plasma Biochemistry in Carp (*Cyprinus carpio L.*). *Acta Vet. Brno*, 71: 117-123
- Malloy HT & Evelyn KA 1937. The determination of bilirubin with the photometric colorimeter. *J. Biol. Chem.*, 119: 481-490.

- Misra L & Wagner H 2007. Extraction of bioactive principles from *Mucuna pruriens* seeds *Indian J. Biochem. & Biophys.*, 44(1): 56-60.
- Navarro MC, Montilla MP, Jiménez J, Martín A & Utrilla MP 1993. Free radical scavenger and antihepatotoxic activity of *Rosmarinus tomentosus*. *Planta Med.*, 59: 312-314.
- Nermeen AM, Hassan OK, Helala AE & Sayed M 2013. Hepatoprotective and Antioxidant Effects of Commiphora Against CCl₄ Induced Liver Injury in Adult Male Albino Rats. *Int. J. Toxicol. & Pharmacol. Res.*, 5(4):79-86.
- Odebiyi OO & Sofowora EA 1978. Phytochemical screening of Nigerian medicinal plants II. *Lloydia*, 41(3): 234-246.
- Omojasola PF & Awe S 2004. The Antibacterial activity of the leaf extract of *Anacardium occidentale* and *Gossypium hirsutum* against some selected microorganisms. *Biosci. Res. Commun.*, 16(1).
- Plummer DT 1978: In: An Introduction to Practical Biochemistry. 2nd ed. McGraw-Hill, London, pp 144-145.
- Oyagade JO, Awotoye OO, Adewumi JT & Thorpe HT 1999. Antimicrobial activity of some Nigeria medicinal plants. *Biosci. Res. Commun.*, 11(3): 193-197.
- Pourgholam R, Soltani M, Hassan DM, Ghoroghi A, Nahavandi R & Pourgholam H 2006. Determination of diazinon LC50 in grass carp (*Ctenopharyngodon idella*) and the effect of sub-lethal concentration of toxin on some hematological and biochemical indices. *Iranian J Fish Sci.*, 5: 67-82.
- Reitman S & Frankel S 1957. A colorimetric method for the determination of serum glutamic oxalacetic and glutamic pyruvic transaminases. *Am. J. Clin. Pathol.*, 28(1): 56-63
- Rodkey FI 1965. Direct Spectrophotometric Determination of Albumin in Human Serum. *Clin Chem.*, 11: 478-487.
- Rolls GO 2008. *An Introduction to Specimen Preparation*. Accessed 15 Aug. 2016. www.leicabiosystems.com/pathologyleaders/introduction-to-specimen-preparation
- Sathiyarayanan L & Arulmonzhi S 2007. *Mucuna pruriens*: A comprehensive review. *Pharmacognosy Review*, 1(1): 157-162.
- Scriver CR, Beaudet AL, Sly WS, Valle D, Stanbury JB, Wyngaarden JB, Fredrickson & DS Eds. 2000. *The Metabolic and Molecular Basis of Inherited Disease*. McGraw-Hill Professional, New York, p. 6338.
- Seifert WF, Bosma A, Hendricks HF, Roholl PJ VanLeeuwen RE, VanThiel-DeRuiter GC, Seifert-Bock I & Knook DL 1994. Vitamin A deficiency potentiates carbon tetrachloride-induced liver fibrosis in rats. *Hepatology*, 19(1): 193-201.
- Shahjahan M, Sabitha KE, Jamu, M & Shyamala-Devi CS 2004. Effect of *Solanum trilobatum* against carbon tetrachloride induced hepatic damage in albino rats. *Indian J. Med. Res.*, 120: 194-198.
- Sharma K, Dutta N, Pattanaik AK & Hasan QZ 2003. Replacement value of undecorticated sunflower meal as a supplement for milk production by crossbred cows and buffaloes in the Northern plains of India. *Trop. Anim. Health Prod.*, 35: 131-145
- Sherlock S & Dooley J 2008. *Diseases of the liver and biliary systems*. John Wiley and Sons, New Jersey, p. 728.
- Spencer JPE, Jenner A, Butler J, Aruoma OI, Dexter DT, Jenner P, & Halliwell B 1996. Evaluation of the prooxidant and antioxidant actions of L-Dopa and dopamine in vitro: Implications for Parkinson's disease. *Free Radical Res.*, 24: 95-105.
- Spencer JPE, Jenner P & Halliwell B 1995. Superoxide-dependent GSH depletion by L-Dopa and dopamine: Relevance to Parkinson's disease. *Neuroreport*, 6(1): 1480-1484.
- Spina MB & Cohen G 1988. Exposure of school synaptosomes to L-Dopa increases levels of oxidised glutathione. *J. Pharmac. & Experimental Therapeutics*, 247(1): 502-507.
- Tackett AJ, DeGrasse JA, Sekedat MD, Oeffinger M, Rout MP & Chait BT 2005. I-DIRT, a general method for distinguishing between specific and nonspecific protein interactions. *J. Proteome Res.*, 4(5): 1752-1756.
- VanDeursen VM, Damman K, Hillege HL, VanBeek AP, VanVeldhuisen DJ & Voors AA 2010. Abnormal liver function in relation to hemodynamic profile in heart failure patients. *J. Cardiac Failure*, 16(1): 84-90.
- Vicenová B, Vopálenký V, Burýsek L & Pospíšek M 2009. Emerging role of interleukin-1 in cardiovascular diseases. *Physiol. Res.*, 58(4): 481-98.
- Wennberg RP 2000. The blood-brain barrier and bilirubin encephalopathy. *Cellular & Molecular Neurobio.*, 20(1): 97-109.
- Wu J, Dannielson A & Zern M 1999. Toxicity of hepatotoxins: New insights into mechanisms and therapy. *Expert Opinion On Investig. Drugs*, 8(5): 585 – 607.
- Yakubu MO, Taiwo IA & Otabor JI 2016. Genotoxicity and reproductive effects of chronic consumption of aqueous bitter leaf extract in albino rats. *Fuw Trends Sci. & Techn. J.*, 1: 202-205.